

AMENDMENTS TO THE SPECIFICATION

In the written description:

Please amend the paragraph in the present specification beginning on page 28, line 24 and ending on page 29, line 17 as follows:

Bone marrow cells were collected from B6 mouse thighbone and tibia. Then these bone marrow cells were separated by using NycoPrep to obtain bone marrow mononuclear cells. These bone marrow mononuclear cells were cultured in a Petri dish for 10 to 12 hours and unbound bone marrow mononuclear cells were collected. The unbound bone marrow mononuclear cells were mixed with magnetic bead-labeled c-kit antibody and cultured. Next, c-kit-positive cells were separated with the use of a magnetic cell sorter (MACS; manufactured by Miltenyi Biotec). These c-kit-positive cells were cultured at a concentration of ~~2.5x10⁵~~ 2.5x10⁵ cells/ml in the presence of SCF (provided by Dr. Sudo, Toray), Flt3L (manufactured by Immunex), GM-CSF (4 ng/ml) and IL-4 (10 ng/ml) for 7 to 9 days to give bone marrow-derived dendritic cell precursors. ~~These c-kit positive cells were cultured at a concentration of 2.5x10⁵ cells/ml in the presence of SCF (provided by Dr. Sudo, Toray), Flt3L (manufactured by Immunex), GM-CSF (4 ng/ml) and IL-4 (10 ng/ml) for 7 to 9 days to give bone marrow-derived dendritic cell precursors.~~

Please amend the paragraph beginning on page 30, line 7 and ending on page 31, line 11 as follows:

CD3-positive T lymphocytes were prepared from B6 mouse spleen cells by using a magnetic cell sorter (MACS; manufactured by Miltenyi Biotec). Then these T lymphocytes were mixed with the antigen-primed dendritic cell precursors (the bone marrow-derived dendritic cell precursors having been primed with the B16 cancer cell line lysate) to give a ratio of 1:20 and cultured in a 24-well plate in the presence of IL-2 and IL-7. After culturing the T lymphocytes for 5 days, the culture was continued while replacing 50% of the medium with a fresh one at intervals of 2 or 3 days. On the days 7 and 14, antigen-primed dendritic cell precursors were newly added to thereby re-prime the T lymphocyte under culturing. On the day 21 of the culture, the T lymphocytes were collected. Subsequently, the T lymphocytes having been cultured were mixed with the peripheral blood-derived dendritic cell precursors (derived from the mice administered with dead *P. acnes* cells or purified MIP-1 α) having been primed with the B16 cancer cell line lysate, the bone-marrow-derived dendritic cell precursors having been primed with the MMC-treated B16 cancer cell line lysate, the dendritic cell precursors having been primed with an EL4 cancer cell line lysate, unprimed dendritic cell precursors or the cancer cell line lysate alone in a 96-well round bottomed plate (manufactured by Nunc) and cultured at 37°C for 4 to 5 days. After the completion of the culture, 15 μ l/well of a 5 mg/ml MMT (~~3-(4,5-dimethylthiazolyl-2-yl)-2,5-diphenyltetrazolium bromide~~) (3-(4,5-dimethylthiazolyl-2-yl)-2,5-diphenyltetrazolium bromide) was added and the culture was continued at 37°C for additional 4 hours. After the completion of the culture, the absorbance of the contents in each well was measured at 550 nm, thereby measuring the proliferation of the T lymphocyte.

Please amend the paragraph beginning on page 32, line 1 and ending on page 32, line 15 as follows:

B6 mice were divided into groups (each having 8 animals) and subcutaneously injected in the abdominal side with the peripheral blood-derived dendritic cell precursors (obtained from the mice administered with dead *P. acnes* cells or purified MIP-1 α) having been primed with the B16 cancer cell line lysate, the bone marrow-derived dendritic cell precursors having been primed with the B16 cancer cell line lysate, the dendritic cell precursors having been primed with the EL4 cancer cell line lysate, unprimed dendritic cell precursors (~~$1 \times 10^6/\text{animal in each case}$~~), ($1 \times 10^6/\text{animal in each case}$), the cancer cell line lysate alone or PBS. After 14 days, B16 cancer cells (~~$2 \times 10^5/\text{animal}$~~) ($2 \times 10^5/\text{animal}$) were subcutaneously inoculated into abdominal side of the mice. Following the inoculation, the tumor area was measured at intervals of 3 days to judge the proliferation of the tumor. Simultaneously, surviving mice were counted.

Please amend the paragraph beginning on page 34, line 14 and ending on page 35, line 15 as follows:

<7> Spleen cells were collected from B6 mice showing disappearance of cancer cells on the day 60 after the B16 cancer cell line inoculation from among the B6 mice immunized with the peripheral blood-derived dendritic cell precursors (obtained from the mice administered with MIP-1 α) having been primed the B16 cancer cell line lysate and the B6 mice immunized with the bone marrow-derived dendritic cell precursors in the above <5>. CD8-positive T lymphocytes (1×10^6) were separated from the spleen cells and cultured together with MMC-treated B16 cancer cell line (~~4×10^5~~) (1×10^5) for 5 days. Then the cultured T lymphocytes were serially diluted and added to a 96-well plate at a ratio of 100 $\mu\text{l}/\text{well}$. Next, a cell suspension of the B16 cancer cell line was added at a ratio of 100 $\mu\text{l}/\text{well}$. After culturing at 37°C for 10 hours, the plate was centrifuged and 100 $\mu\text{l}/\text{well}$ of the supernatant was transferred into a fresh 96-well plate. Then a reaction solution in a cytotoxicity detection kit (LDH; manufactured by Boehringer

Mannheim) was added to each well at a ratio of 100 μ l/well. After reacting the reaction plate at room temperature for 30 minutes, the absorbance thereof was measured at 490 nm with a plate absorption meter to thereby assay the cytotoxic activity. In an experiment on MHC class I restriction inhibition, the B16 cancer cells were treated with anti-MHC class I antibody (anti-H2Kb/H2Db antibody) or a control antibody (anti-H2Ds antibody) at 37°C for 30 minutes before the addition.

Please amend the paragraph beginning on page 35, line 19 and ending on page 36, line 13 as follows:

<8> On the days 0 and 7, B6 mice were subcutaneously immunized in the abdominal side with the peripheral blood-derived dendritic cell precursors (obtained from the mice administered with dead *P. acnes* cells or purified MIP-1 α) having been primed with the B16 cancer cell line lysate, the bone marrow-derived dendritic cell precursors having been primed with the B16 cancer cell line lysate, the dendritic cell precursors having been primed with an EL4 cancer cell line lysate, unprimed dendritic cell precursors (~~1x10⁶/animal in each case~~), (1x10⁶/animal in each case), the cancer cell line lysate alone or PBS. Seven days after the second immunization, spleen cells were collected from the immunized mice. Then, CD4-positive cells and CD8-positive cells, i.e. T cells, were separated from these spleen cells by using a cell sorter. These T cells were mixed with MMC-treated B16 cancer cells and cultured by using a 24-well plate for 48 hours. Next, the cytotoxic activity was assayed with the use of the cells thus cultured. Further, the IFN γ concentration in the supernatant of the cultured cells was measured by using an IFN γ ELISA system (manufactured by Endogen).

Please amend the paragraph beginning on page 36, line 15 and ending on page 37, line 2 as follows:

B16 cancer cells (~~$1 \times 10^6/\text{animal}$~~) ($1 \times 10^6/\text{animal}$), were injected into B6 mice via the tail vein. On the days 3 and 7 following the injection of the cancer cells, the peripheral blood-derived dendritic cell precursors (obtained from the mice administered with MIP-1 α) having been primed with the B16 cancer cell line lysate, the bone marrow-derived dendritic cell precursors having been primed with the B16 cancer cell line lysate, unprimed dendritic cell precursors ($1 \times 10^6/\text{animal}$ in each case), the cancer cell line lysate alone or PBS were each injected into the mice via the tail vein. On the day 21 following the cancer cell injection, mouse lungs were surgically taken out and metastatic foci were counted, thereby judging the degree of metastasis.